

Effects of preoperative nutritional support on colonic anastomotic healing in malnourished rats

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ABSTRACT

Objective: It has been proven that malnutrition increases postoperative morbidity and mortality, and it may also negatively affect wound healing in the gastrointestinal tract. In the literature, there is only one study evaluating the effects of preoperative nutritional support on colonic anastomotic healing under malnourished conditions. In order to improve the data on this topic, an experimental study was planned to evaluate the effects of preoperative nutritional support on colonic anastomotic healing in malnourished rats.

Material and Methods: The study included 18 male Wistar albino rats divided into 3 groups. The control (C) group was fed ad libitum for 21 days. The malnutrition (M) group and preoperative nutrition (P) group were given 50% of the daily food consumed by the rats in Group C for 21 days to induce malnutrition. At the end of 21 days, Group P was fed ad libitum for 7 days (preoperative nutritional support). Colonic transection and end-to-end anastomosis was performed at 21 days in Group C and Group M and at 28 days in Group P. The rats were sacrificed at postoperative 4 days, anastomotic bursting pressure was measured, and samples were taken to analyze tissue hydroxyproline levels.

Results: Anastomotic bursting pressure was significantly higher in Group C than in Group M and Group P ($p<0.05$), and it was significantly higher in Group P than in Group M ($p<0.05$). Tissue hydroxyproline levels in Group P were found to be significantly higher than those in Group M and Group C ($p<0.05$).

Conclusion: One week of preoperative nutritional support increases collagen synthesis in the colon and positively affects anastomotic healing under malnourished conditions.

Keywords: Malnutrition, nutritional support, anastomotic healing

INTRODUCTION

It has been shown that malnutrition increases postoperative morbidity and mortality and prolongs the length of hospital stay in patients undergoing surgical interventions (1-4). According to the recent data, there are approximately 30 million individuals with malnutrition or nutritional risk in Europe, and it is estimated that €170 billion is spent annually for this purpose (5). The effects of malnutrition on various organ systems and wound healing have been evaluated in experimental and clinical studies; particularly, its effects on the gastrointestinal system have been emphasized (3, 6-8). Some clinical studies have indicated lower general complication rates by preoperative nutritional support in patients with malnutrition undergoing major gastrointestinal surgery (9-11). However, there is little experimental research on this topic.

Multiple local and systemic factors affect colonic anastomotic healing. To our knowledge, there is only one experimental study evaluating the effects of preoperative nutritional support on colonic anastomotic healing under malnourished conditions in the literature (12). In order to increase the knowledge on this subject, an experimental study assessing the effects of preoperative nutritional support on colonic anastomotic healing in malnutrition-induced rats was planned.

MATERIAL AND METHODS

Experimental Model

The study was approved by the Animal Ethics Committee of Ankara Hospital. Eighteen male Wistar albino rats weighing approximately 210 g were included. Test subjects, housed under a 12-h light:dark cycle, were divided into 3 groups (Figure 1): malnutrition (M) group ($n=6$); preoperative nutrition (P) group ($n=6$), and control (C) group ($n=6$). The rats were kept in separate cages throughout the experiment, and their weights were measured and recorded daily.

Group C was fed ad libitum for 21 days. Group M and Group P were given 50% of the daily food consumed by the rats in Group C for 21 days. Thereafter, Group P was fed ad libitum (preoperative nutrition) for 7 days.

After 21 days, the rats in Group M and Group C were prepared for surgery. They were deprived of food for 12 h before the intervention, their skin was shaved, the surgical site was sterilized using 10% povidone-iodine, and laparotomy was performed through a 3-cm midline incision under general anesthesia

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[45 mg/kg ketamine (Eczacıbaşı, Istanbul, Turkey) and 5 mg/kg IM xylazine (Alfasan, Woerden, Netherlands)]. The caecum was identified and a 1-cm colon segment was resected 5 cm distal to the ileocecal valve, and single-layer end-to-end anastomosis was performed using 8 sutures of 4-0 prolene sutures. The abdominal layers were closed with 2-0 silk sutures, and both groups were fed ad libitum after surgery. At postoperative day 4, re-laparotomy was performed under general anesthesia. An 18 G catheter was placed 2 cm proximal to the anastomotic line. The colon was ligated with a 2-0 silk tie 2 cm distal to the anastomotic line and over the catheter. A special apparatus was used to measure the bursting pressure. The colon segment was inflated using 0.9% NaCl at 5 ml/min. The pressure values were followed-up from the monitor; a steep fall or a plateau on the curve was considered as the anastomotic bursting pressure (ABP) and recorded. After that, the rats were sacrificed using intracardiac blood aspiration. Keeping the anastomotic line in the middle, a 2-cm colon segment was resected for the measurement of tissue hydroxyproline (Hyp) levels, the sutures were removed, and the tissue was maintained at -80°C for biochemical analysis.

The test subjects in Group P also underwent the same procedures at 28 days.

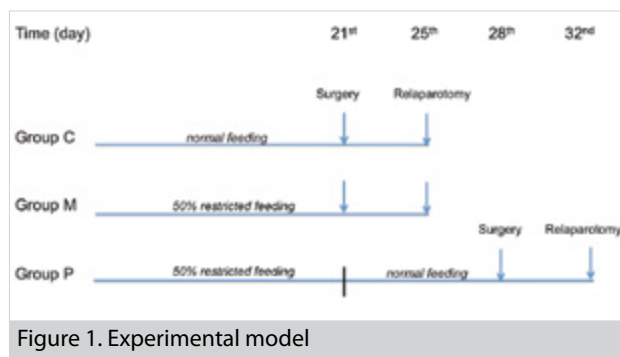
Biochemical Methods

Hyp standards (2–20 μg) were prepared from the stock solution. The total volume of the standards and samples was adjusted to 50 μL with sodium hydroxide. The dry weight of the samples was measured and they were hydrolyzed by autoclaving at 120°C for 20 min. Chloramine T (450 μL) (Kimetsan, Istanbul, Turkey) was added to the hydrolyzate and the mixture was stirred slowly. In order to provide oxidation, the samples were kept at room temperature for 25 min. Ehrlich's reagent (500 μL) was added to each sample, they were mixed, and the chromophore was developed after incubation for 20 min at 65°C . The absorbance of the reddish purple mixtures was read with a spectrophotometer (Jenway, Stafford, UK) at 550 nm and the values were presented as mg/g tissue (13).

Statistical Analysis

The numerical parameters were presented as mean, standard deviation, and minimum and maximum values. The statistical significance level was set at $p=0.05$. Statistical analysis was performed using SPSS 16.0 program (SPSS Inc., Chicago, IL, USA).

The Kruskal–Wallis test was used to compare the 3 groups. Paired comparison of the groups was performed using the Mann–Whitney U test; intra-group comparisons were made using the Wilcoxon test.



The effects of tissue Hyp levels on ABP were evaluated by linear regression analysis.

Spearman's correlation test was performed to determine the correlation of the preoperative weights of the subjects with ABP and tissue Hyp levels and the correlation of tissue Hyp levels with ABP.

RESULTS

None of the 18 rats in the study died and the study was completed with 18 rats. There was no significant difference between the groups in terms of the baseline weight ($p=0.077$). After 21 days of restricted feeding, the mean weight loss in Group M and Group P was 21.8% and 23.8%, respectively, compared with the baseline weight. There was no significant difference between the 2 groups in terms of weight loss ($p=0.317$). In addition, the mean weight (in grams) in Group P and Group M were similar ($p=0.417$). After 21 days, the mean weight in Group C showed a significant difference from that in Group M and Group P ($p=0.004$).

Seven days of preoperative nutrition after 21 days of restricted feeding resulted in 20% weight increase (199.5 ± 9.25 g) in Group P, and there was a significant difference between Group M and Group P in terms of the mean preoperative weight ($p=0.012$). In addition, there was a statistically significantly difference between Group P and Group C in terms of the preoperative weight ($p=0.004$). Weight alterations observed in the study groups during the study are summarized in Table 1.

In re-laparotomy, during dissection of adhesences, anastomotic separation developed in a rat in Group C. Complete anastomotic separation was observed in a rat in Group M, although the site was bounded by the omentum and the bowel. In a rat in Group P, 50% anastomotic separation was present and a plug of stool obstructed the distal portion of the anastomosis.

There was a significant difference between the 3 groups in terms of ABP. ABP in Group P was significantly higher than that in Group M ($p=0.009$) and lower than that in Group C ($p=0.047$). ABP in Group M was lower than that in Group P and Group C ($p=0.009$) (Table 2).

A significant difference was found between the 3 groups in terms of tissue Hyp levels ($p=0.001$). Tissue Hyp levels in Group P were significantly higher than those in Group M ($p=0.004$) and Group C ($p=0.016$). Tissue Hyp levels in Group M were lower than those in both Group P ($p=0.004$) and Group C ($p=0.008$) (Table 2).

The statistical comparison of the groups in terms of ABP and tissue Hyp levels are summarized in Figure 2.

In regression analysis, 1 unit increase in tissue Hyp levels yielded 5.246 units of ABP increase in Group M, 1.513 units of increase in Group P, and 9.118 units of increase in Group C. While 15.2% of changes in ABP could be explained by tissue Hyp levels in Group M, 19.5% and 48.1% of changes in ABP could be explained by tissue Hyp levels in Group P and Group C, respectively. However, this association was not statistically significant.

Correlation analysis showed that there was a moderate positive correlation between the preoperative body weight and

Table 1. Weight (g) alterations in the study groups during the experiment

	Group C	Group M	Group P
Baseline	206.0±4.98 (200–213)	216.5±13.4 (194–230)	219.8±9.26 (208–229)
21 days	275.0±30.21 (231–311)	169.33±14.36 (152–195)	165.5±7.06 (160–179)
Surgery	275.0±30.21 (231–311)	169.33±14.36 (152–195)	199.5±9.25 (175–215)

Table 2. Tissue hydroxyproline levels and anastomotic bursting pressure according to the study groups

Parameter	Group M	Group C	Group P
Hydroxyproline (µg/mg)	1.7±0.4	3.0±0.6	4.8±1.4
Bursting pressure (mm-Hg)	26.8±5.1	48.2±3.8	40.0±5.2

ABP. With the increase in the preoperative weight, ABP was found to be increased ($r=0.520$). This relation was statistically significant ($p=0.027$).

There was a moderate positive correlation between the preoperative weight and tissue Hyp levels. Tissue Hyp levels increased with the increase in the preoperative weight ($r=0.444$). However, this relationship was not statistically significant ($p=0.065$).

There was a significant moderate positive correlation between tissue Hyp levels and ABP ($p=0.020$). ABP increased with the increase in tissue Hyp levels ($r=0.542$).

DISCUSSION

Intestinal anastomotic healing is affected by various variables such as the primary disease, local blood flow, presence of infection, surgical technique, and nutrition status. Healing in colonic anastomosis is a more complex and slow process than that in the other regions of the digestive system. Currently, animal experiments are widely used to understand this complicated mechanism. Although colonic anastomosis is technically difficult in rats, rats were used in this research as they are easy to maintain and care for and suitable for the planned model with rapid wound healing.

There are various studies assessing the effects of malnutrition on colonic anastomotic and wound healing in the literature (6-8, 12, 14). These studies have numerous discrepancies such as in terms of the type and degree of malnutrition. Although protein malnutrition is generally induced in these studies, the parameters affecting the degree of malnutrition, such as the duration and degree of protein restriction, are different. This situation makes it difficult to compare the obtained data and to reach a common opinion. In our study, we used balanced protein-energy malnutrition by restricting all nutrients, as defined by Gonçalves et al. (12). Because induction of malnutrition was long (21 days) in this model, severe catabolic processes were avoided and therefore the loss of test subjects could be kept at minimum.

ABP and tensile strength are widely used in estimating the strength of colonic anastomosis; however, it is debatable as to which should be used as the gold standard. An experimental

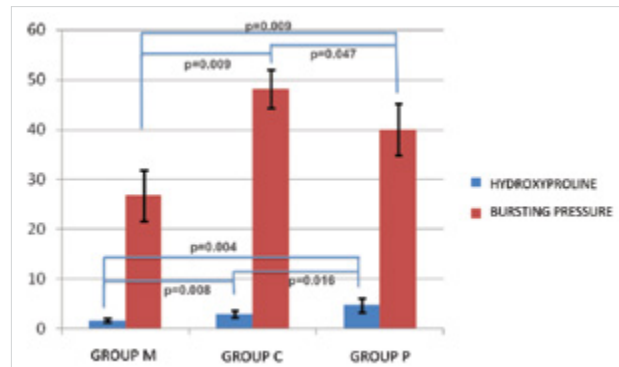


Figure 2. Comparison of anastomotic bursting pressure and tissue hydroxyproline levels in the study groups

study involving 48 rats compared ABP and tensile strength and found no significant difference between them on postoperative day 4 (15). Law et al. (8) showed that rupture occurred somewhere outside the anastomotic line at bursting pressure measurements on postoperative day 4. Therefore, we assessed anastomotic strength on postoperative day 4 in this study. To minimize the probable measurement errors and application differences during assessment, a pressure monitor and an infusion pump were used. Thus, the results obtained in this study can be compared with those in further experimental studies.

The main mechanisms providing wound healing are the amount and nature of collagen released at the wound site. The most widely used method in estimating the amount of synthesized collagen is the detection of tissue Hyp levels. Another method that can be used is histological evaluation using fluorescent staining. However, as this method is expensive, difficult to perform, and requires special equipment and experience, tissue Hyp level measurement was decided to be used in this present study.

In the previous studies on nutritional status and intestinal wound healing, the effect of preoperative nutritional support was not evaluated. In the study by Irvin et al. (6), induction of severe protein malnutrition (weight loss: 34%) in rats was shown to negatively affect colonic anastomotic healing. In the study by Mukerjee et al. (16) in rats fed without protein supplementation, colonic tensile strength was found to be low. Ward et al. (7) included 3 groups of rats in their study. The animals in the first and second groups were fed a low-protein diet, and 12.5% weight loss, which was considered mild malnutrition, was induced. The first group was fed a normal diet after surgery, whereas the low-protein diet was continued in the second group. The animals in the third group were fed a normal diet throughout the experiment. ABP in the first and second groups (that had negative nitrogen balance) was lower than that in the control group, and early postoperative normal feeding was shown to improve this situation. No group received preoperative nutritional support in these studies.

In the study performed by Law et al. (8), colonic anastomosis was performed in rats fed a low-protein diet for 2 weeks to achieve 12% weight loss, and ABP and tissue Hyp levels were measured at days 4 and 7. Although the ABP in the experimental group was lower than that in the control group, there was no difference in tissue Hyp levels. Irvin et al. (6) determined colonic weight loss in malnourished rats and reported that this decrease was predominantly due to the loss of the non-

collagen part. The authors suggested that this caused a relative increase in collagen in the colon. The higher tissue Hyp levels in Group P when compared with those in Group C and Group M in our study may be similarly explained by the relative increase in collagen.

The relation between preoperative nutritional support and anastomotic healing was first evaluated by Gonçalves et al. (12) in 2009. That study used a protocol similar to that used in the present study and found significantly higher anastomotic tensile strength in the group that received preoperative nutritional support than that in the malnourished group. There was no difference compared with the control group. The collagen levels in the preoperative nutritional support group were significantly higher than those in the malnourished group and similar to those in the control group. We achieved similar degrees of weight loss and malnutrition in our study. There was significant weight loss in Group P and Group M compared with the baseline weight and compared with Group C at 21 days. No difference was found between Group P and Group M in terms of weight loss and malnutrition rates. These findings indicate the success of the method used in the induction of malnutrition. There was 20% weight increase in Group P after 7 days of preoperative nutritional support. Weight increase was significantly higher than that in Group M. This finding proves the efficacy of preoperative nutritional support. When the mean preoperative body weight (in grams) in Group P and Group C are considered, it is seen that preoperative nutritional support is effective; however, it is remarkable that body weight values could reach the values obtained in Group C. According to the current nutrition guidelines, the aim of preoperative nutritional support is not to regain all the weight lost due to chronic malnutrition but to cease weight loss and, if possible, move to the positive side; the main aim is to decrease the negative effects of malnutrition on surgical results (17).

In our study, there was a statistically significant difference between the 3 groups in terms of ABP measurements. ABP in Group P was significantly higher than that in Group M, whereas it was lower than that in Group C. However, achieving values close to the mean values obtained in Group C suggest that preoperative nutritional support positively contributes to anastomotic strength. Significantly lower values obtained in Group M compared with both Group P and Group C supports this implication.

The effect of tissue Hyp levels on ABP was evaluated using regression analysis. Tissue Hyp levels had a certain degree of effect on ABP; however, this effect was different between the groups. Data obtained from regression analysis suggest that tissue Hyp levels had an effect on anastomotic healing; however, they were not the main factor determining anastomotic strength. The limitation of the effect is consistent with the opinion that the anastomotic healing process is affected by numerous independent variables. Correlation analysis was used to test the relationship and the interaction between tissue Hyp levels and ABP. According to the analysis, a significant moderate positive correlation was found between tissue Hyp levels and ABP. This finding suggests that anastomotic strength increases parallel to the increase in collagen synthesis in the wound. This is a predictable result. There was a significant moderate positive correlation between the preoperative weight of the test subjects and ABP. Recovery of a part of the

body weight by nutritional support had a positive effect on anastomotic strength in malnourished subjects.

A test subject in Group C developed iatrogenic anastomotic separation during dissection. Complete anastomotic separation was observed in a test subject in Group M. A test subject in Group P developed 50% separation at the anastomotic line and the distal portion of the anastomosis was obstructed with stool. These subjects were excluded during ABP measurements. However, samples were taken from the anastomotic sites to compare anastomotic strength and tissue Hyp levels. Tissue Hyp levels of the 2 subjects with anastomotic separation in Group C and Group M were lower than those of the other test subjects in these groups. Tissue Hyp levels of the subject in Group P was above the mean values. These findings also indicate that there is a positive relationship between anastomotic strength and collagen synthesis. Thus, it can be accepted that anastomotic separation that occurred in the subject in Group P was due to the stool plug at the distal portion.

CONCLUSION

According to the results of this study, it was concluded that preoperative nutritional support under malnourished conditions has a positive effect on colonic anastomotic healing and collagen synthesis. Evaluation of the preoperative weight, tissue Hyp levels, and ABP and the relationship between these 3 parameters indicated that the wound healing process has a highly variable nature. In malnutrition, every step of this process is negatively affected. Providing efficient and correct nutritional support is important in minimizing the negative effects of malnutrition on colonic anastomosis. However, in order to elucidate the complex process of intestinal wound healing, other variables should also be experimentally tested.

Ethics Committee Approval: Ethics committee approval was received for this study from the ethics committee of Animal Ethics Committee of Ankara Hospital.

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REFERENCES

1. Malone DL, Genuit T, Tracy JK, Gannon C, Napolitano LM. Surgical site infections: reanalysis of risk factors. *J Surg Res* 2002; 103: 89-95. [\[CrossRef\]](#)
2. Correia MI, Waitzberg DL. The impact of malnutrition on morbidity, mortality, length of hospital stay and costs evaluated through a multivariate model analysis. *Clin Nutr* 2003; 22: 235-239. [\[CrossRef\]](#)
3. Sungurtekin H, Sungurtekin U, Balci C, Zencir M, Erdem E. The influence of nutritional status on complications after major intraabdominal surgery. *J Am Coll Nutr* 2004; 23: 227-232. [\[CrossRef\]](#)
4. Schiesser M, Müller S, Kirchhoff P, Breitenstein S, Schäfer M, Clavien PA. Assessment of a novel screening score for nutritional risk in predicting complications in gastro-intestinal surgery. *Clin Nutr* 2008; 27: 565-570. [\[CrossRef\]](#)

5. Ljungqvist O, De Man F. Under nutrition: a major health problem in Europe. *Nutr Hosp* 2009; 24: 368-370.
6. Irvin TT, Hunt TK. Effect of malnutrition on colonic healing. *Ann Surg* 1974; 180: 765-772. [\[CrossRef\]](#)
7. Ward MW, Danzi M, Lewin MR, Rennie MJ, Clark CG. The effects of subclinical malnutrition and refeeding on the healing of experimental colonic anastomoses. *Br J Surg* 1982; 69: 308-310. [\[CrossRef\]](#)
8. Law NW, Ellis H. Revised model for the study of colonic anastomotic healing in protein malnourished rats. *Eur Surg Res* 1989; 21: 218-223. [\[CrossRef\]](#)
9. Detsky AS, Baker JP, O'Rourke K, Goel V. Perioperative parenteral nutrition: a meta-analysis. *Ann Intern Med* 1987; 107: 195-203. [\[CrossRef\]](#)
10. Von Meyenfeldt MF, Meijerink WJ, Rouflart MM, Builmaassen MT, Soeters PB. Perioperative nutritional support: a randomised clinical trial. *Clin Nutr* 1992; 11: 180-186. [\[CrossRef\]](#)
11. Klein S, Kinney J, Jeejeebhoy K, Alpers D, Hellerstein M, Murray M, et al. Nutrition support in clinical practice: review of published data and recommendations for future research directions. National Institutes of Health, American Society for Parenteral and Enteral Nutrition, and American Society for Clinical Nutrition. *JPEN* 1997; 21: 133-156. [\[CrossRef\]](#)
12. Gonçalves CG, Groth AK, Ferreira M, Matias JE, Coelho JC, Campos AC. Influence of pre-operative feeding on the healing of colonic anastomoses in malnourished rats. *JPEN* 2009; 33: 83-89. [\[CrossRef\]](#)
13. Reddy GK, Enwemeka CS. A simplified method for the analysis of hydroxyproline in biological tissues. *Clin Biochem* 1996; 29: 225-229. [\[CrossRef\]](#)
14. Zaizen Y, Ford EG, Costin G, Atkinson JB. Stimulation of wound bursting strength during protein malnutrition. *J Surg Res* 1990; 49: 333-336. [\[CrossRef\]](#)
15. Ikeuchi D, Onodera H, Aung T, Kan S, Kawamoto K, Imamura M, et al. Correlation of tensile strength with bursting pressure in the evaluation of intestinal anastomosis. *Dig Surg* 1999; 16: 478-485. [\[CrossRef\]](#)
16. Mukerjee P, Mepham JA, Wapnick S, Datta BN, Cox AG. The effect of protein deprivation on alimentary healing. *J Surg Res* 1969; 9: 283-288. [\[CrossRef\]](#)
17. Weimann A, Braga M, Harsanyi L, Laviano A, Ljungqvist O, Soeters P, et al. ESPEN guidelines on enteral nutrition: surgery including organ transplantation. *Clin Nutr* 2006; 25: 224-244. [\[CrossRef\]](#)