



Does taurolidine have any effect on liver regeneration and oxidation in the experimental hepatectomy model?

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ABSTRACT

Objective: Taurolidine is a bicyclic molecule produced by the natural amino acid taurine. Antibacterial, antiendotoxic and cytoprotective effects of taurolidine have been shown experimentally. Data on the effects of taurolidine on oxidative stress and hepatic regeneration are limited. The aim of the study was to evaluate the effect of taurolidine on hepatic regeneration and oxidative stress in rats undergoing partial hepatectomy.

Material and Methods: Forty adult, male Wistar Albino rats were randomly divided into four equal groups: sham (S) group (n= 10), post-sham operation taurolidine administered (ST) group (n= 10), partial hepatectomy (H) group (n= 10) and post-partial hepatectomy taurolidine administered (HT) group (n= 10). 100 mg/kg/day taurolidine was administered for seven days. Blood and liver tissue samples were collected on postoperative day seven. Liver tissue malondialdehyde, glutathione and Cu-Zn superoxide dismutase activity (SOD) were measured to assess oxidative stress. Binuclear hepatocyte and Ki-67 antigen levels were measured to evaluate hepatic regeneration.

Results: There was no difference between the groups for malondialdehyde, Cu-Zn superoxide dismutase and glutathione levels ($p > 0.05$). Binuclear nuclei levels were comparable between the H and HT groups ($p = 0.06$), while taurolidine decreased binuclear hepatocyte levels in the sham operated groups ($p = 0.02$). Taurolidine application decreased Ki-67 levels after partial hepatectomy ($p = 0.001$).

Conclusion: Taurolidine may cause anti-regenerative effects after partial hepatectomy without causing oxidative damage.

Keywords: Oxidative stress, liver resection, hepatic regeneration, partial hepatectomy

INTRODUCTION

Partial hepatectomy (PH) is a common procedure of liver surgery. Hepatic regeneration (HR), which almost instantly follows PH, is an exceptional response of the organ to recover to its original size and functions (1). HR is a complex process where epithelial cells, Kupffer cells and hepatocytes have significant roles (2). Remarkable biochemical changes take place during HR; the oxidative stress has a paramount role with an immediate increase after the PH and during hepatic regeneration (3-4). Moreover, free oxygen radicals arise from the partial fragmentation of molecular oxygen, and free radicals increase with tissue damage; tissue malondialdehyde (MDA) is utilized to quantify lipid peroxidation caused by free radicals (5,6). Finally, tissue glutathione (GSH) levels and superoxide dismutase (SOD) activities reveal antioxidant status (7,8).

Taurolidine (TRD) is a taurine based bicyclic molecule, and previous studies have revealed that it has antibacterial, antiendotoxic and cytoprotective effects (9,10). TRD is mostly used for the treatment of patients with sepsis. It leads to prokaryote degeneration and connects free lipopolysaccharides to exotoxins (11). TRD suppresses mitochondrial cytochrome-C and induces apoptosis in tumor cells by inhibiting protein synthesis at an early stage without adverse effects (12,13). Since there is limited data on the effects of TRD on oxidative damage and hepatic HR after PH, it was aimed to evaluate this issue in an animal model.

MATERIAL and METHODS

The Institutional Ethics Committee of Experimental Animal Research approved the protocol of the study. Forty Wistar-Albino male rats, weighing between 250 and 350 gr, were used. Rats were housed five in per cage, under constant temperature (

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20–22°C) and humidity (50F) with an artificial 12-hour light-dark cycle and randomized into four groups. Rats were fed ad libitum with standardized industrial rat prey during the study period except 12 hours prior to operation. Rats were divided randomly into four equal weight-matched groups containing ten rats each: Group 1: Sham (S) group; Group 2: Subsequent to sham operation, intraperitoneal TRD administered (ST) group; Group 3: Partial hepatectomy (H) group (the control group); Group 4: Subsequent to partial hepatectomy, intraperitoneal TRD administered (HT) group.

A daily dose of 100-mg/kg/day (Tauroline[®], Boehringer Ingelheim, Germany) were given to the rats in the ST and HT groups. The S and H groups were administered same volume 0.9% NaCl solution intraperitoneally daily, while the others received TRD injections.

PH was performed with general anesthesia. After shaving, the abdominal wall was cleaned with povidone iodine solution and a median abdominal incision was performed. Standard 70% PH was performed in the H and the HT groups (14), whereas laparotomy without liver resection was performed in the S and the ST groups. The abdominal incision was closed with 2/0 silk continuous sutures. On postoperative day seven, the rats were decapitated under general anesthesia [ketamine 40 mg/kg, intraperitoneal and xylazine (5 mg/kg, intramuscular)]. Liver tissue sampling was done immediately after decapitation. Oxidative stress was evaluated by measuring malondialdehyde, glutathione and Cu-Zn superoxide dismutase activity (SOD) levels in the liver tissue.

Biochemical analyses

Tissue samples were weighed at room temperature. Ten percent homogenates were prepared by homogenization buffer with 0.15 M phosphate. Homogenization was performed at 4°C. When the homogenization was completed, supernatants were obtained by centrifuging the homogenates, which were prepared for the determination of MDA at 3000 rpm for 10 minutes and for SOD and GSH at 13000 rpm for 15 minutes. The method of Beuter and et al. (15) was used for these quantifications. SOD analysis was performed based on Sun and Oberley's method (16). In MDA analysis, a pink colored complex is formed as a result of its incubation at 95°C with thio-barbituric acid (TBA) at pH 3.5 and this complex is measured by spectrophotometer at a wavelength of 532 nm. MDA levels were given as nmol/gr tissue by using MDA's molar extinction coefficient ($n = 1.56 \times 10^5 \text{ M}^{-1}$).

Histopathological analysis

For each animal, four randomly taken tissue sections were stained with hematoxylin-eosin and examined by experienced pathologists who were unaware of the treatment groups. Dual nuclei hepatocytes located randomly selected ten different fields were counted under x400 magnification. Cells definitely defined that two nuclei in a single cytoplasm were accepted as dual nuclei and the arithmetic average of the total number of cells were considered. The number of mitosis located in the same fields was recorded.

Immunohistochemical staining procedure was applied with Ki-67 (SP6) (Neomarkes, USA) ready-to-use rabbit monoclonal antibody. Ki-67 positive hepatocyte nuclei were counted at randomly selected 10 different fields under x400 magnification on aforementioned laminas. Some cells, such as lymphocyte, Kupffer et cetera, were excluded from the count. The ratio of Ki-67 positive nuclear staining cells to total cells number was calculated by percentage (17).

Statistics

The results were presented as mean and standard deviation. P value of <0.05 was considered to be significant used by JMP[®] Statistical Discovery Software and analyzing Kruskal-Wallis test. Student's t test was used in the comparison of the groups of Binuclear hepatocyte and Ki-67.

RESULTS

All the rats completed the experiment; there was no perioperative death. There was no difference between the groups regarding the malondialdehyde, Cu-Zn superoxide dismutase and glutathione levels (Table 1). On postoperative day seven, both experimental and control groups had similar hepatic oxidant activity.

Although they were significantly close, the levels of binuclear hepatocytes were statistically comparable between the H and HT groups. ($p = 0.06$). Additionally, taurolidine administration decreased binuclear hepatocyte numbers in the sham-operated group (Figure 1) and (Table 2). Further evaluation with immunohistochemical staining revealed decreased Ki-67 levels after PH with taurolidine use (Figure 2) and (Table 2).

Table 1. Groups' oxidative stress parameters levels

	Malondialdehyde $\mu \text{ mol/wet tissue}$	Superoxide Dismutase U/wet tissue	Glutathione $\mu \text{ mol/wet tissue}$
Sham (n= 10)	12.6 \pm 7.7	0.9 \pm 0.3	1.5 \pm 0.3
Sham and taurolidine (n= 10)	12.8 \pm 5.2	0.9 \pm 0.1	1.3 \pm 0.4
Partial hepatectomy (n= 10)	5.9 \pm 0.7	0.8 \pm 0.2	1.3 \pm 0.2
Partial hepatectomy and taurolidine (n= 10)	6.02 \pm 1.8	0.9 \pm 0.2	1.5 \pm 0.5
p	0.96	0.38	0.28

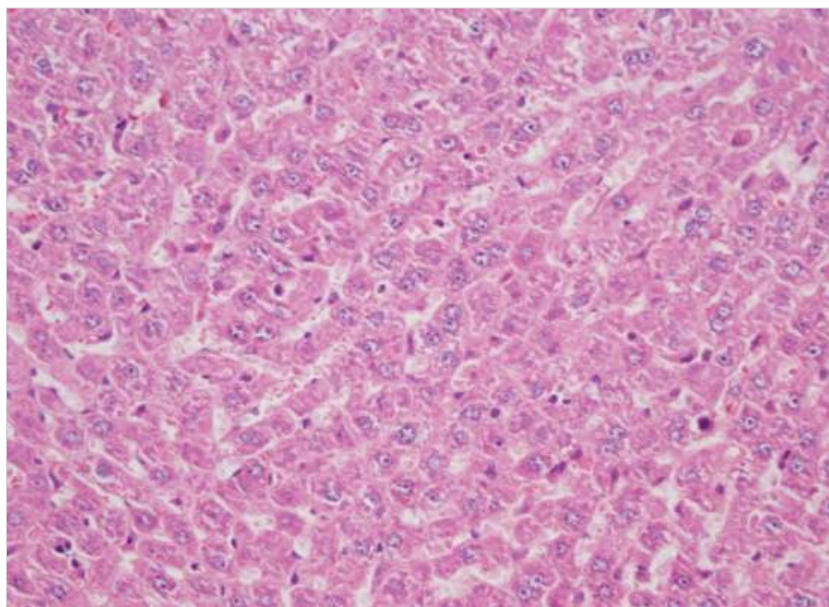


Figure 1. Binuclear hepatocytes after taurolidine administration.

Table 2. Binuclear and Ki-67 levels in groups		
	Binuclear hepatocyte	Ki-67 (median)
Sham (n= 10)	15.6 ± 2.5	1
Sham and taurolidine (n= 10)	12.4 ± 2.3	1
p*	0.002	1
Partial hepatectomy (n= 10)	9.2 ± 2.2	10.3
Partial hepatectomy and taurolidine (n= 10)	11.2 ± 1.9	4.03
p**	0.06	0.003

* p between the groups of sham with sham and taurolidine.
 ** p between partial hepatectomy with partial hepatectomy and taurolidine.

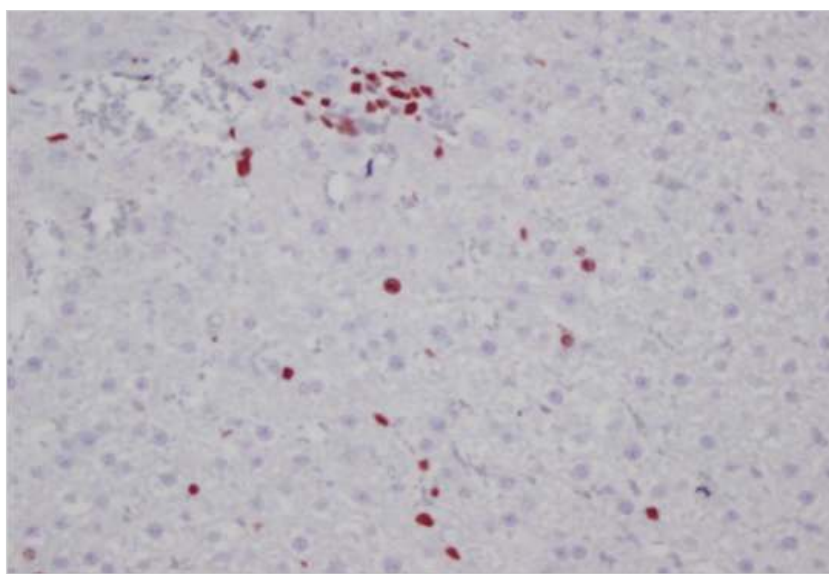


Figure 2. Ki-67 levels after taurolidine administration.

DISCUSSION

The results of the current study revealed that TRD use may reduce the ability of *in vivo* regeneration of the liver. This is a remarkable finding to consider for use of TRD after PH, especially for patients with limited functional liver capacity. TRD is commonly used for treatment of patients with sepsis (11). While we did not use any kind of septic model in our experimental study, it is well known that the toxic internal environment limits regenerative capacity of the liver and cause hepatotoxicity (18). The additional anti-proliferative effect of TRD may be an undesirable effect following liver resection. Anti-neoplastic effects of TRD against various cancer types such as mesothelioma have been shown previously (19). Prooxidant antineoplastic drugs affect cancerous tissue, but do not affect non-neoplastic tissue (20). TRD is a prooxidant antineoplastic drug with anti-regenerative effect on cancer cells and not hazardous on non-neoplastic tissues (21). TRD promotes leukopoiesis, erythropoiesis, as well as improves renal and hepatic function against systemic toxicity in severe sepsis (7). While TRD supports wound healing, it may prolong the epithelization process (22,23). Since regeneration pathways varies for different, consequences of TRD use may vary among the tissue types (24).

Hepatic regeneration is an exceptional response of the liver to recover to its original size and functions (1). In addition to hepatocytes, Epithelial cells, endothelial and Kupffer cells of the biliary tract play a role in liver regeneration (1,2). Endocrine and paracrine hormones, proto-oncogenes and transcription factors are the key points for modulating regeneration process (25). Remarkable biochemical changes take place during HR; the oxidative stress has a paramount role with an immediate increase after the PH and during hepatic regeneration (3,4). The liver tissue oxidant status was similar among study groups. The oxidant activity increases within the early phase of liver regeneration (26). Regeneration starts in 10-12 hours after hepatectomy, reaches to its maximum value at 24 hour and completes in 5-10 days (27). In rats, lipid peroxidation significantly increases 90 minutes after the generation of the liver ischemia (28). Hepatic functional tests are closely correlated with the MDA (29). Therefore, monitoring lipid peroxidation, gives a reliable impression on liver functions. We have previously reported that high oxidant activity was being maintained in the plasma but resolved in the liver tissue seven days after PH (14). Overall impression based on the results of the study is that anti-regenerative effects of TRD after PH seem non-toxic. There is no sign of oxidative damage in the biochemical and histological analyses were observed.

We need to mention the controversies on oxidative activity related to *in vivo* and *in vitro* use of TRD. Dual effect of TRD has been shown previously. While antioxidant properties of TRD

have been shown against sepsis induced oxidative stress, TRD increased production of reactive oxygen species in *ex vivo* glioma cells (7-21). Low production of oxidants and dominant antioxidant capacity of the non-malignant cells compared to the cancer cells and infectious agents may establish a selective environment for survival of healthy host tissues (30).

It has been shown that many inflammatory cytokines including TNF-alpha and IL-6 are up-regulated during hepatic regeneration (31,32). TRD increases NFkB which inhibits NFkB in esophageal cancer (33). NFkB is known as potential regulator of TNF-alpha and IL-6. Anti-inflammatory effects of TRD may indirectly contribute inhibition of hepatic regeneration (34). Angiogenesis is an important component of liver regeneration (35). Tumor-derived VEGF are blocked by TRD (29). Antiangiogenic properties of TRD can be considered as a potential drawback for hepatic regeneration.

Formaldehyde, which is a carcinogenic and toxic substance, is an *in vivo* breakdown product of TRD. In routine practice, TRD is used with polyvinylpyrrolidone, which reduces *in vivo* concentration of formaldehyde (36). No toxic effect of TRD has been reported neither in clinical practice nor in our study (37). In addition to hematoxylin and eosin dye for histological evaluation, immunohistochemical staining with Ki-67 (SP6) had performed. Ki-67 is an antigen in the nucleus of the cell and the level of the monoclonal antibody, which is produced against Ki-67, increases as the cell cycle progresses on (38,39). Ki-67 analysis is one of the most reliable immunohistochemical techniques for evaluating liver regeneration (40).

Lack of molecular markers analyzing hepatic regeneration and oxidative stress is the limitation of the study. However, our results provide a basis on taurolidine use after PH or in liver surgery for the patients with limited hepatic parenchyma reserve.

CONCLUSION

Taurolidine may cause anti-regenerative effects after partial hepatectomy without causing oxidative damage.

Ethics Committee Approval: The approval for this study was obtained from Istanbul University Animal Experiments Local Ethics Committee (Decision No: 41, Date: 16.04.2009).

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REFERENCES

- Bedirli A, Kerem M, Pasaoglu H, Erdem O, Ofluoglu E, Sakrak O. Effects of ischemic preconditioning on regenerative capacity of hepatocyte in the ischemically damaged rat livers. *J Surg Res* 2005; 125(1): 42-8. [\[CrossRef\]](#)
- Michalopoulos GK. Liver regeneration after partial hepatectomy: critical analysis of mechanistic dilemmas. *Am J Pathol* 2010; 176(1): 2-13. [\[CrossRef\]](#)
- Contreras-Zentella ML, Hernandez-Munoz R. Is liver enzyme release really associated with cell necrosis induced by oxidant stress? *Oxid Med Cell Longev* 2016; 2016: 3529149. [\[CrossRef\]](#)
- Carnovale CE, Scapini C, Alvarez ML, Favre C, Monti J, Carrillo MC. Nitric oxide release and enhancement of lipid peroxidation in the regenerating rat liver. *J Hepatol* 2000; 32(5): 798-04. [\[CrossRef\]](#)
- Reilly PM, Schiller W, Bulkley GB. Pharmacologic approach to tissue injury mediated by free radicals and other reactive oxygen metabolites. *Am J Surg* 1991; 161(4): 488-03. [\[CrossRef\]](#)
- Olinescu R, Bardoc R, Militaru M, Nita S, Dimitriu L. The changes of peroxidase and total antioxidant in the plasma of patients who received I131 therapeutically. *Rev Roum Med Int* 1992; 30(2): 113-17. [\[CrossRef\]](#)
- Han D, Hanawa N, Saberi B, Kaplowitz N. Mechanism of liver injury. III. role of glutathione redox status in liver injury. *Am J Gastrointest Liver Physion* 2006; 291(1): G1-7. [\[CrossRef\]](#)
- Singh M, Sandhir R, Kiran R. Effects on antioxidant status of liver following atrazine exposure and its attenuation by vitamin E. *Exp Tox Pathol* 2011; 63(3): 269-76. [\[CrossRef\]](#)
- Akkus A, Gulmen M, Cevik A, Bildik N, Sad O, Oztürk E, et al. Effect of peritoneal lavage with taurolidine on primary colonic anastomosis in a rat model of secondary peritonitis. *Surg Today* 2006; 36(5): 436-40. [\[CrossRef\]](#)
- Doddakula KK, Neary PM, Wang JH, Sookhai S, O'Donnell A, Aherne T, et al. The antiendotoxin agent taurolidine potentially reduces ischemia/reperfusion injury through its metabolite taurine. *Surgery* 2010; 148(3): 567-72. [\[CrossRef\]](#)
- Liu Y, Zhang A, Cao L, Xia HT, Mia JJ. Taurolidine lock solutions for the prevention of catheter related bloodstream infections: a systematic review and meta-analysis of randomized controlled trials. *PLoS One* 2013; 8(11): e79417. [\[CrossRef\]](#)
- Braumann C, Stuhlder B, Bobrich E, Menenakos C, Rogalla S, Jacobi CA. High doses of taurolidine inhibit advanced intraperitoneal tumor growth in rats. *J Surg Res* 2005; 129(1): 129-35. [\[CrossRef\]](#)
- Braumann C, Menenakos C, Atanasou V, Pfirrmann RW, Guenther N, Jacobi CA. Leukopoiesis is not affected after intravenous treatment with the novel antineoplastic agent Taurolidine. *Eur Surg Res* 2008; 40(4): 341-46. [\[CrossRef\]](#)
- Saribeyoglu K, Aytac E, Pekmezci S, Saygili S, Uzun H, Ozbay G, et al. Effects of clonitilolite treatment on oxidative stress after partial hepatectomy in rats. *Asian J Surg* 2011; 34(4): 153-7. [\[CrossRef\]](#)
- Beuter E. Glutathione. In: *Red Cell Metabolism A Manual of Biochemical Methods*. 2nd ed. New York: Grune and Stratton, 1975: 112-14. [\[CrossRef\]](#)
- Sun Y, Oberley LW. Suitability of copper chloride as a reaction terminator for superoxide dismutase activity assay. *Clin Chim Acta* 1994; 226(1): 101-03. [\[CrossRef\]](#)
- Ngala Kenda JF, Hemptinne B, Lambotte L. Role of metabolic overload in the initiation of DNA synthesis following partial hepatectomy in the rat. *Eur Surg Res* 1984; 16(5): 294-9. [\[CrossRef\]](#)
- Xu CPI, Liu J, Liu JC, Han DW, Zhang Y, Zhao YC. Dynamic changes and mechanism of intestinal endotoxemia in partially hepatectomized rats. *W J Gastroenterol* 2007; 13(26): 3592-97. [\[CrossRef\]](#)
- Aceto N, Bertino P, Barbone D, Tassi G, Manzo L, Porta C, et al. Taurolidine and oxidative stress: a rationale for local treatment of mesothelioma. *Eur Respir J* 2009; 34(6): 1399-407. [\[CrossRef\]](#)
- Raj L, Ide T, Gurkar AU, Foley M, Schenone M, Li X, et al. Selective killing of cancer cells by a small molecule targeting the stress response to ROS. *Nature* 2011; 475(7355): 231-4. [\[CrossRef\]](#)
- Rodak R, Kubota H, Ishihara H, Eugster HP, Könü D, Möhler H, et al. Induction of reactive oxygen intermediates-dependent programmed cell death in human malignant ex vivo glioma cells and inhibition of the vascular endothelial growth factor production by taurolidine. *J Neurosurg* 2005; 102(6): 1055-68. [\[CrossRef\]](#)
- Presterl E, Schahawi MD, Lusignani LS, Paula H, Reilly J. Basic principles and introduction to disinfectants and antiseptics for skin, mucosa and wounds. *Basic Microb and Inf* 2018; 10: 3; 52-56. [\[CrossRef\]](#)
- Fahrner R, Möller A, Press AT, Kortgen A, Kiehnopf, Rauchfuss F, et al. Short-term treatment with taurolidine is associated with liver injury. *BMC Pharmacol Toxicol* 2017; 18(61): 1-9. [\[CrossRef\]](#)
- Simpson JM, Séguin B, Gitelman AI. Effects of topical application of taurolidine on second intention healing of experimentally induced wounds in rats. *Am J Vet Res* 2008; 69(9): 1210-16. [\[CrossRef\]](#)
- Sica A, Invernizzi, Mantovani A. Macrophage plasticity and polarization in liver homeostasis and pathology. *Hepatol* 2014; 59(5): 2034-42. [\[CrossRef\]](#)
- Tao Y, Wang M, Chen E, Tang H. Liver regeneration: analysis of the main relevant signaling molecules. *Med Inf* 2017; 1-9. [\[CrossRef\]](#)
- Min JS, Deangelis RA, Reis ES, Gupta S, Maurya MR, Evans C, et al. Systems analysis of the complement-induced priming phase of liver regeneration. *J Immunol* 2016; 197(6): 2500-08. [\[CrossRef\]](#)
- Wang L, Wang X, Xie G, Wang L, Hill CK, Deleve LD. Liver sinusoidal endothelial cell progenitor cells promote liver regeneration in rats. *J Clin Invest* 2012; 122(4): 1567-73. [\[CrossRef\]](#)
- Ayan F, Aytac E, Saygili S, Genç H, Bayrak I, Uzun H, et al. Effects of lycopene on oxidative stress and remnant liver histology after partial hepatectomy in rats. *Turk J Gastroenterol* 2011; 22(4): 408-13. [\[CrossRef\]](#)
- Wondrak GT. Redox-directed cancer therapeutics: molecular mechanisms and opportunities. *Antioxid Redox Signal* 2009; 11(12): 3013-69. [\[CrossRef\]](#)
- Fujiyoshi M, Ozaki M. Molecular mechanism of liver regeneration and protection for treatment of liver dysfunction and diseases. *J Hep Bil Panc Sci* 2011; 18(1): 13-22. [\[CrossRef\]](#)
- Blindenbacher A, Wang X, Langer I, Savino R, Terracciano L, Heim MH. Interleukin 6 is important for survival after partial hepatectomy in mice. *Hepatology* 2003; 38(3): 674-82. [\[CrossRef\]](#)
- Daigeler A, Chromik AM, Geisler A, Bulut D, Hilgert C, Krieg A, et al. Synergistic apoptotic effects of taurolidine and TRAIL on squamous carcinoma cells of the esophagus. *Int J Oncol* 2008; 32(6): 1205-20. [\[CrossRef\]](#)

34. Marcinkiewicz J, Kurnyta M, Biedroń R, Bobek M, Kontny E, Maśliński W. Anti-inflammatory effects of taurine derivatives (taurine chloramine, taurine bromamine, and taurolidine) are mediated by different mechanisms. *Adv Exp Med Biol* 2006; 583: 481-92. [CrossRef]
35. Drixler TA, Vogten MJ, Ritchie ED, van Vroonhoven TJ, Gebbink MF, Volest EE, et al. Liver regeneration is an angiogenesis-associated phenomenon. *Ann Surg* 2002; 236(6): 703-11. [CrossRef]
36. Braumann C, Tangermann J, Jacobi C, Mueller J, Dubiel W. Novel anti-angiogenic compounds for application in tumor therapy-COP9 signalosome-associated kinases as possible targets. *Mini Rev Med Chem* 2008; 8(5): 421-28. [CrossRef]
37. Gong L, Greenberg HE, Perhach JL, Waldman SA, Kraft WK. The Pharmacokinetics of taurolidine metabolites in healthy volunteers. *Clin Pharmacol* 2007; 47(6): 697-703. [CrossRef]
38. Gerdes J, Schwab U, Lemke H, Stein H. Production of a mouse monoclonal antibody reacts with a human antigen associated with cell proliferation. *Int J Cancer* 1983; 31(1): 13-20. [CrossRef]
39. Gerdes J, Lemke H, Barsch H, Wacker HH, Schwab U, Stein H. Cell cycle analysis of a cell proliferation associated human nuclear antigen defined by the monoclonal antibody Ki-67. *J Immunology* 1984; 133(4): 1710-5. [CrossRef]
40. Assy N, Minuk GY. Liver regeneration methods for monitoring and their applications *J Hepatol* 1997; 26(4): 945-52. [CrossRef]



ORJİNAL ÇALIŞMA-ÖZET

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Taurolidinin deneysel hepatektomi modelinde hepatik rejenerasyon ve oksidatif stres üzerine herhangi bir etkisi var mıdır?

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ÖZET

Giriş ve Amaç: Taurolidin doğal bir aminoasit olan taurinden oluşan bisiklik yapıda bir moleküldür. Taurolidinin antibakteriyel, antiendotoksik ve sitoprotektif etkileri deneysel çalışmalarla gösterilmiştir. Taurolidinin oksidatif stres ve hepatik rejenerasyona etkileri için ise elimizde yeterli veri bulunmamaktadır. Çalışmanın amacı parsiyel hepatektomi yapılan ratlarda taurolidinin hepatik rejenerasyon ve oksidatif stres üzerine etkilerini ölçmektir.

Gereç ve Yöntem: Kırk erkek Wistar Albino rat rastgele dört eşit gruba ayrıldı: sham (S) grubu (n= 10), sham operasyonu sonrası taurolidin (ST) uygulanan grup (n= 10), parsiyel hepatektomi (H) grubu (n= 10), parsiyel hepatektomi sonrası taurolidin (HT) uygulanan grup (n= 10) 100 mg/kg/gün taurolidin yedi gün boyunca uygulandı. Kan ve karaciğer doku örnekleri postoperatif yedinci günden sonra alındı. Oksidatif stresi saptamak amacıyla karaciğer dokusunda malonaldehit, glutatyon ve Cu-Zn superoksit dismutaz (SOD) aktivitesi ölçüldü. Binükleer hepatosit ve Ki-67 anti-jen seviyeleri hepatik rejenerasyonu değerlendirmek için ölçüldü.

Bulgular: Gruplar arasında malonaldehit, Cu-Zn superoksit dismutaz ve glutatyon seviyeleri açısından fark yok (p> 0,05). Binükleer nükleus seviyeleri H ve HT grupları arasında karşılaştırılabilir düzeydeyken (p= 0,06), taurolidin binükleer hepatosit seviyelerini sham ve sham opere gruplarda azalttı. Taurolidin uygulaması parsiyel hepatektomi sonrası Ki-67 seviyelerini düşürdü (p= 0,001).

Sonuç: Taurolidin parsiyel hepatektomi sonrası oksidatif hasara yol açmadan anti-rejeneratif etki gösteriyor olabilir.

Anahtar Kelimeler: Oksidatif stres, karaciğer rezeksiyonu, hepatik rejenerasyon, parsiyel hepatektomi

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